

Neurochemical Dysregulation of Covid 19 Infection

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Introduction

Coronaviruses are so named because of the halo of spike (S) proteins that decorate their surfaces [1,2]. These S proteins interact with specific cellular receptors to bind host cells and the binding is followed by protease-mediated S protein cleavage, which exposes fusion-promoting domains that enable viral entry. SARS-CoV-2 infects cells through interactions between its S protein and the Angiotensin I Converting Enzyme 2 (ACE2) receptor on target cells. ACE2 plays a crucial modulatory role in the renin-angiotensin system, which regulates blood pressure, salt and water balance [3]. Infection requires S protein cleavage, likely by the host cell serine protease TMPRSS2 (Transmembrane protease, serine 2) although other proteases may also be involved. SARS-CoV-2 belongs (Severe acute respiratory syndrome-related coronavirus 2) to the coronavirus family, which includes the pandemic MERS-CoV (Middle East respiratory syndrome coronavirus) and SARS-CoV (SARS (Severe acute respiratory syndrome)-associated coronavirus) and the lesser known but more common endemic coronaviruses HCoV-OC43 (*Human coronavirus OC43*), HCoV-HKU1, HCoV-229E, and HCoV-NL63. The endemic coronaviruses can infect the upper airway and frequently cause the common cold, which in turn is associated with both acute and chronic changes in smell and taste [4,5]. The main proposed mechanisms for acute viral-mediated changes in smell include conductive deficits caused by loss of patency due to swelling of the mucosa and increased mucus production, changes in mucus composition, and secondary changes in olfactory signaling caused by local release of inflammatory intermediates like cytokines. While cold-causing viruses likely act through multiple mechanisms to influence smell, recovery from virus-associated olfactory deficits tend to resolve with a time course similar to that of other cold-related symptoms like nasal congestion. With the exception of HCoV-NL63, the endemic coronaviruses do not use ACE2 as their primary cellular receptor [6], a molecular distinction that likely underlies key differences in pathophysiology. SARS-CoV also uses ACE2 as its main receptor, and in one case study SARS-CoV infection was

associated with anosmia [7], although (unlike COVID-19) chemosensory disturbances are not a hallmark of SARS. Differences between SARS-CoV and SARS-CoV-2 in terms of their impacts on chemosensory systems may relate to biophysical differences, as the receptor-binding domain of the SARS-CoV-2 spike protein binds ACE2 with higher affinity and with a different binding mode than that of SARS-CoV [8]. Species variation in the protein sequence of ACE2 significantly affects its affinity for the SARS-CoV-2 S protein, which in turn renders distinct model organisms differentially susceptible to infection [9]. Given data suggesting that ACE2 is necessary for SARS-CoV-2 to infect host cells, researchers have used a variety of approaches to discern the pattern of expression of ACE2 and other viral entry proteins across the tissue landscape, with the goal of inferring possible target cells and disease mechanisms. For example, two studies have reported that cells in the nasal Respiratory Epithelium (RE) have higher expression of SARS-CoV-2 entry genes than cells in the RE that line the trachea or lungs [10]. Consistent with this finding, recent work in macaques, ferrets, and cats identifies the nasal epithelium as a major source of viral RNA after SARS-CoV-2 infection [11]. These data suggest that the nasal epithelium may act as a major reservoir for the virus and SARS-CoV-2 attacks the olfactory system through mechanisms distinct from those used by the more benign endemic coronaviruses. Many patients report anosmia as their first symptom, or in the absence of rhinorrhea or nasal congestion, suggesting that if inflammation is a key component of pathogenesis, it is local rather than generalized [12]; indeed, the sensitivity of anosmia as a predictor of COVID-19 increases in patients without other nasal symptoms [13]. The OE is a complex chemosensory tissue composed of multiple cell types, including immature and mature OSNs (Olfactory sensory neurons), non-neuronal cell types such as sustentacular, Bowman's gland, and microvillar cells, and stem cells including globose and horizontal basal cells. Sustentacular cells are particularly intimately associated with OSNs, and they "enwrap" the sensory dendritic cilia that project into the airspace and enable odor detection [14]. Four recently pub-

lished studies-using species ranging from mouse to human-have explored the cell types in the OE (olfactory epithelium) that express ACE2 and other viral entry genes [15], all four conclude that OSNs do not express ACE2. Instead, co-expression of ACE2 and TMPRSS2 was observed in key support cells (including sustentacular, Bowman's gland, and microvillar cells) and stem cells that repopulate the epithelium after damage. Although ACE2 mRNA was identified using single-cell RNA sequencing (sc-Seq) techniques in only a small subset of these cells, a high level expression of ACE2 protein in a large population of sustentacular cells concentrated in the dorso-medial aspect of the mouse OE, which corresponds to the dorsal "zone" traditionally identified via molecular markers [16]. Consistent with this observation, mouse and human sustentacular cells identified as ACE2 positive by scSeq largely derive from the dorsal zone [17]. The co-expression of ACE2 and TMPRSS2 suggests that OE support cells may be the initial targets of SARS-CoV-2 infection. These inference-based conclusions are increasingly being pressure-tested by experiments in which model organisms are directly subject to SARS-CoV-2 infection. One recent paper in the golden hamster reports that SARS-CoV-2 infects sustentacular cells but not OSNs [18]. Intriguingly, this phenotype is accompanied by damage to the OSN ciliary layer and an increase in the number of microglia present in the OE, both of which are partially reverted to normal at 14 days after infection. A similar study in the hamster identified a large number of cells in the OE that were infected by SARS-CoV-2, although OE cell types were not definitively identified; inspection of the photomicrographs in that paper reveals that most SARS-CoV-2-positive cells traverse the thickness of the OE, suggesting that sustentacular cells are primary targets for infection [19]. Human OE samples obtained from COVID-19 patients have similarly been queried for infection by SARS-CoV-2, which has revealed coronaviral antigens present in OE cells; the infected cell types were not unambiguously identified, but their shape and position is consistent with the virus targeting sustentacular cells rather than OSNs [20]. What mechanisms might link infection of support cells to the acute changes in smell reported in COVID-19. Localized inflammation in the epithelium might block the olfactory clefts, a pair of narrow passages located in the superior regions of the nasal epithelium through which air must flow to reach the OE, which comprises only 5% of the total nasal epithelium in humans [21]. Consistent with this possibility, a recent CT study of a COVID-19 patient who presented with anosmia revealed the blocked olfactory clefts [22]. Alternatively, infection of support cells and the attendant inflammation may cause local increases in inflammatory intermediates such as cytokines, which have been shown to influence OSN function in a non-cell-autonomous manner [23]. Indeed, a recent study demonstrates elevated levels of inflammatory cytokines in the OE of infected patients [24]. Inflammatory intermediates have been suggested to indirectly lower the expression of Odorant Receptor (OR) genes by OSNs, which could cause significant changes in odor perception and this work also shows that OR expression levels return to normal after cessation of the inflammatory insult [25]. Finally, SARS-CoV-2 infection of support cells might alter the OE micro-environment in a manner deleterious to function. Bowman's glands, for example, secrete mucus, which is essential to normal odor detection that bulb neurons, do not express detectable levels of ACE2. Consistent with this finding, targeted deep sequencing of dopaminergic juxtaglomerular cells, which receive direct inputs from nose OSNs, failed to reveal Ace2 mRNA in vascular pericytes in the OB (olfactory bulb) expressed high levels of ACE2 protein, consistent with the reported expression

of ACE2 in perivascular cells in the brain and throughout the body [26]. Pericytes are critical for maintaining the blood-brain barrier, for defining local blood pressure, and for mediating neuroimmune responses [27] and the infection of these cells has the potential to alter perfusion or recruit inflammation, both of which can indirectly influence the function of neural circuits. SARS-CoV-2 can directly infect neurons or glia and it is important to recognize the key caveats in interpreting data suggesting SARS-CoV-2 does or does not infect neurons and glia directly. SARS-CoV-2 in principle has access to the brain through routes independent of the nasal epithelium, including via vasculature and nerves innervating infected tissues; type II and III TRCs (taste receptor cells) in mice reveal that ACE2 is expressed by sour-sensing type III TRCs, and to a lesser extent by bitter and sweet/umami-sensing type II TRCs [28]. While type II and type III TRCs express little to no TMPRSS2, Cathepsins (CTSB, CTSL) are abundant and may function as proteases to cleave SARS-CoV-2 spike protein at type II and III TRCs, type I cells degrade ATP, which is used as a neurotransmitter by type II cells to convey taste information to the brain via the gustatory nerves [29]. If type I cells are indeed ACE2 positive and targeted by SARS-CoV-2, the loss of support cells could lead to taste bud collapse via cell death and/or reduced efficacy of taste signals to gustatory nerves (unlike OSNs-TRCs are not neurons; thus, all of the cell types identified as ACE2 positive in the tongue to date are either stem or epithelial cells. Chemesthetic stimuli are detected by a variety of epithelial sensors and relayed to the brainstem via trigeminal ganglion sensory neurons, distinct branches of which innervate the nasal respiratory epithelium and the non-taste epithelium of the tongue, among other targets [30]. These trigeminal somatosensory afferents are enriched for TRP (transient receptor potential) channel receptors that detect chemical irritants. Chemical perception due to COVID-19 has only just begun, and much remains to be learned about the pathophysiology of SARS-CoV-2 and the current evidence favors a model in which SARS-CoV-2 cell entry genes in the olfactory, gustatory, and chemesthetic systems are not expressed in primary or secondary neurons, but rather are expressed in epithelial, support, and stem cells responsible for maintaining perception. It also remains possible that SARS-CoV-2 directly infects OSNs or bulb neurons. The understanding of molecular mechanisms underlying the chemosensation and the neural pathways that convey information about chemical cues to the brain, still know little about the non-neuronal cells and structures that support sensory transduction. In this sense, SARS-CoV-2 provides an important opportunity to gain insight into how the complex peripheral tissues that support taste, smell, and chemesthesis enable meaningful interactions with the world. Although the evidence that ACE2 is obligate for SARS-CoV-2 entry is strong, it has been suggested that other molecules such as BSG, neuropilin-1, or PIKfyve may participate in SARS-CoV-2 entry [31]. Furthermore, it has recently been reported that low-level expression of ACE2 may be sufficient to support SARS-CoV-2 infection [32,33], suggesting that SARS-CoV-2 may infect apparently ACE2-negative cell types. ACE2 gene is regulated by inflammation in human cells, and other SARS-CoV-2 entry genes may be similarly modulated by primary infection and inflammation [34]. This observation raises the possibility that a broader spectrum of cells expresses ACE2 during SARS-CoV-2 infection than is currently appreciated. A key open question is how primary infection of non-neuronal cells alters chemical perception. Defining the pathophysiological mechanisms underlying anosmia and other SARS-CoV-2-associated disturbances in chemical sensation will require moving past inference based upon gene

or protein expression. In particular, brain chemistry, nociceptive processing, inflammation, and autonomic function are modulated by neurostimulation for different therapeutic purposes.

Coronavirus disease 2019 (COVID-19), which is clinically characterized by fever, myalgia, diarrhea, and respiratory illness, the presentation of patients with extremely low blood oxygenation, but no sensation of dyspnea. Patients admitted with COVID-19 can suffer sudden death after voluntary “breaks” from the oxygen supplementation. Recognizing “happy hypoxia” as a feature of COVID-19 pneumonia has led to better patient care [35], this phenomenon has given rise to the term “happy hypoxemia” has been associated with neurological syndromes such as meningoencephalitis, ischemic stroke, encephalopathy, Guillain-Barré syndrome (GBS), acute necrotizing encephalopathy, and Acute Disseminated Encephalomyelitis (ADEM). Possible mechanisms implicated in these neurological conditions are neuronal injury associated with direct virus infection, hyperinflammation syndrome associated with cytokine storm, a para- or post-infectious inflammatory disease, an immune-mediated disease, or a secondary process due to severe effects of a systemic disorder (sepsis, hyperpyrexia, hypoxia, hypercoagulability, critical illness). Expression of angiotensin converting enzyme 2 (ACE2), the cell entry receptor for SARS-CoV-2, is also distinct between human organs. ACE2 is highly expressed in airway epithelia, kidney cells, small intestine, and lung parenchyma [36], and not surprisingly, this correlates with high viral loads demonstrated in some of these tissues [37]. In the CNS, ACE2 is expressed in neurons, astrocytes, and oligodendrocytes, and it is concentrated in the substantia nigra, ventricles, middle temporal gyrus, posterior cingulate cortex, and the olfactory bulb, although at lower levels than that in lungs [38]. These findings, in addition to the spontaneous recovery of neurological alterations described in most of cases of COVID-19 in the literature support the idea of transient and/or limited SARS-CoV-2 dissemination in the CNS rather than direct damage promoted by SARS-CoV-2 replication. Neurological injury is likely a result of misdirected immune responses either associated with autoimmunity or systemic inflammation in response to limited viral replication, the stimulation of host inflammatory responses involved in subsequent CNS injury, shown by increased levels of total Tau and NfL proteins. Several biomarkers of COVID-19 severity have been identified in blood, including C-reactive protein, D-dimer, lactate dehydrogenase, interleukin-6, and ferritin [39], but less evidence is available for Cerebrospinal Fluid (CSF) biomarkers. Viral antigens have been detected in the brainstem, where the infected regions included the nucleus of the solitary tract and nucleus ambiguus, suggesting that respiratory failure in infected animals or patients may be due to the dysfunction of the cardiorespiratory center in the brainstem. Recently Gordon *et al.* have identified 332 high confidence SARS-CoV-2-human Protein-Protein Interactions (PPIs) and they performed host-virus protein interaction studies by expressing various Open Reading Frames (ORFs) of COVID-19 in HEK293 cells, including 16 nonstructural proteins (Nsp1-16) which form the Replicase/Transcriptase Complex (RTC), four structural proteins: spike (S), envelope (E), membrane (M), and nucleocapsid (N) 17, and nine putative accessory factors [40,41].

Two proteins, the spike protein (S) and main protease Nsp5 are potential molecules as a prominent number of interactions were related to lipid modifications and vesicle trafficking to S protein whereas Nsp5 showed high confidence interaction with the epigenetic regulator histone deacetylase 2 (HDAC2). Both these molecules are potential targets for therapeutic inhibi-

tion as on one hand lipid modulation can promote neuroinvasion and on the other hand, HDAC2 which is highly expressed in neuronal tissues plays a crucial role in CNS development and is a potential therapeutic target of neurodegenerative diseases. There is an urgent need to understand the neurotropic potential of the COVID-19 virus by understanding the possible mechanism of neuromodulation which will be significant for the prevention and treatment of the SARS-CoV-2-induced respiratory failure [42]. In the current scenario, various research strategies need to be employed to understand the mechanisms of neuromodulation resulting either directly in respiratory failure or causing neurological deficits. Host-virus interaction studies using either omics approach [43] (genomic/transcriptomic/proteomic/lipidomic/metabolomic) or cloning and expressing individual COVID-19 proteins in different neuronal cell lines followed by pull-down assays and mass spectrometry and analyzing the interacting protein network. Further proteomic/chemoinformatic analysis can help in identifying already available drugs and clinical molecules from the databases that might interfere with the viral-human interactome. SARS-CoV-1 has been reliably detected in brain tissue specimens of autopsy donors with SARS, specifically in the cytoplasm of neurons in the cortex and hypothalamus, sometimes associated with neuronal edema and nuclear degeneration [44]. Examination of autopsy tissue from a patient with encephalitis revealed neuronal necrosis, glial cell hyperplasia, and infiltration of monocytes and T cells. Additionally, virions were visualized in neurons on electron microscopy, and SARS-CoV-1 RNA was isolated from the specimen. In murine models, SARS-CoV-1 enters the CNS via the olfactory bulb and exhibits rapid transsynaptic spread. Viral neuroinvasion [45] could plausibly be achieved by several routes, including transsynaptic transfer across infected neurons, entry via the olfactory nerve, infection of vascular endothelium, or leukocyte migration across the blood-brain barrier (BBB). There is increasing evidence that human and nonhuman CoV invade peripheral nerve terminals, spread retrograde along nerve synapses, and gain access to the CNS [46]. Transsynaptic transfer of virus has been demonstrated for several CoV, including HCoV-OC43, hemagglutinating encephalomyelitis virus 67 (HEV67), and avian bronchitis virus. For example, HEV67 enters the oronasal cavity and infects the nasal mucosa, lung epithelium, and small intestine of suckling piglets and rats. It then infects the peripheral nerves and spreads retrograde to the dorsal root ganglion, ending in the medullary neurons. A membrane coating-mediated endocytotic or exocytotic pathway facilitates HEV67 transfer between motor cortex neurons [47]. A similar, vesicle-mediated secretory pathway allows HEV67 to spread between neurons and satellite cells. For intracellular spread within a neuron, fast axonal transport uses axonal microtubules to move molecules retrograde or anterograde [48]. Herpes simplex virus, HIV, and HCoV-OC43 have all been shown to use retrograde fast axonal transport to infect the cell body of neurons. The BBB (Blood Brain Barrier) is composed of vascular endothelium, astrocytes, pericytes, and extracellular matrix. Vascular endothelial cells are joined by tight junctions and regulate the permeability of the BBB. Endothelia throughout the body express ACE2 and are at risk for infection by SARS-CoV-2. Once the virus gains access to vascular and neuronal tissue, it could begin a cycle of viral budding and further damage vascular and neuronal tissue as the virus comes into contact with ACE2 on neurons, glia, and vessels [49]. The SARS-CoV-1 virus has been shown to infect lymphocytes, granulocytes, monocyte derivatives, and monocytes, which all express ACE2 [50]. It has been demonstrated that T lymphocytes allow SARS-CoV-2 infection

but do not support viral replication. The systemic inflammation that characterizes COVID-19 likely increases the permeability of the BBB, thereby allowing infected immune cells, cytokines, and possibly virus to pass into the CNS. Severe COVID-19 disease is characterized by increased IL-2, IL-6, IL-7, granulocyte-colony-stimulating factor, interferon- γ inducible protein 10, monocyte chemoattractant protein 1, macrophage inflammatory protein 1- α , and tumor necrosis factor α . and cytokine storm likely contributes significantly to toxic-metabolic encephalopathy in severe cases [51]. For neurochemical evidence of neuronal injury and glial activation in patients with moderate and severe COVID-19, the clinical utility of monitoring plasma NFL and GFAP are investigated. Severe COVID-19 had higher plasma concentrations of GFAP and NFL than controls, while GFAP was also increased in patients with moderate disease. In patients with severe disease, an early peak in plasma GFAP decreased on follow-up, while NFL showed a sustained increase from first to last follow-up, perhaps reflecting a sequence of early astrocytic response and more delayed axonal injury [52]. Plasma biomarkers of CNS injury by single molecule array, neurofilament light chain protein (NFL; a marker of intra-axonal neuronal injury) and glial fibrillary acidic protein (GFAP; a marker of astrocytic activation/injury), nature of COVID-19-related CNS damage and its relation to both clinically defined CNS events such as hypoxic and ischemic events and mechanisms more closely linked to systemic severe acute respiratory syndrome coronavirus 2 infection and consequent immune activation, Increased CSF levels of NFL, GFAP and total-tau protein were seen in 63%, 37%, and 16% of patients, respectively. Increased NFL correlated with disease severity, time in intensive care and level of consciousness. NFL in CSF was higher in patients with central neurological symptoms [53]. Neurological symptoms included altered mental status (42%), headache (42%), central (21%) and peripheral weakness (32%). Minor pleocytosis and increased immunoglobulin G levels in CSF are detected and the neuronal autoantibody testing using commercial tests was negative. SARS-CoV-2 replication in CNS may trigger infiltration by immune cells. COVID-19 patients with neurological diseases present oligoclonal bands in the CSF [54]. Chloroquine and hydroxychloroquine, initially developed as antimalarial drugs, work by preventing the acidification of endosomes, which interrupts cellular functions and may prevent viral entry via ACE2 binding. Hydroxychloroquine inhibits SARS-CoV-2 in vitro, but in vivo studies are lacking [55]. Particularly, the two largest randomized controlled trials to date (RECOVERY and WHO SOLIDARITY), both confirmed that CQ/HCQ regimen does not provide any clinical benefit for COVID-19 patients [56].

Tocilizumab is a monoclonal antibody to the IL-6 receptor that may attenuate cytokine release in patients with severe inflammatory disease. There are limited retrospective data that suggest possible benefit and it has poor penetration into the CNS [57]. Remdesivir is a viral RNA-dependent RNA polymerase inhibitor and in vitro data have shown that it is a potent SARS-CoV-2 inhibitor [58-60].

Molecular docking technique is used to analyze the binding of non-structural proteins to porphyrin. The porphyrin in the human body is mainly iron porphyrin, namely heme and many hemes are not free but bound to hemoglobin. Viruses have a massive demand for porphyrins [61]. Therefore, the novel coronavirus may target hemoglobin, attack heme, and acquire porphyrin. In order to study the attack behavior of nsp16-nsp10 (orf1ab), ORF3a, and ORF10 proteins, used ZDOCK molecular docking technology to examine these three proteins and it can

analyze protein interactions and find the approximate location of these three proteins on hemoglobin. For oxidized hemoglobin, nsp16-nsp10 acted on the middle bottom of the alpha and beta chain and closed to the alpha chain. ORF3a acted at the bottom of the beta chain and ORF10 acted below the alpha chain. The possible mechanism was that nsp16-nsp10 first attacked the alpha chain, and then, ORF3 and ORF10 successively attack the beta chain. For deoxyhemoglobin, nsp16-nsp10 acted on the top of the 1-beta. ORF3a acted at the bottom of the 1-beta. ORF10 acted on the top of 1-beta and ORF3 and ORF10 have embedded in deoxyhemoglobin and directly docked to the heme of the beta chain and it indicates that the viral protein can attack heme on hemoglobin. The possible mechanism was that nsp16-nsp10 first attacked the 1-beta chain, and then, ORF3 and ORF10 successively attack the 1-beta. It is challenging to perform molecular simulations. Due to the close distance of the attack postures of some proteins, so it is unclear the order of three proteins attacked. The nsp16-nsp10 molecule may be an essential protein, playing a vital role throughout the attack. It is worth noting that the above simulation shows that the deoxyhemoglobin is more vulnerable than oxidized hemoglobin. Attack of oxidized hemoglobin by viral proteins will lead to less and less hemoglobin that can carry oxygen, carbon dioxide and blood sugar. People with diabetes can have unstable blood sugar and body cells have extreme inflammation due to excess iron, carbon dioxide and oxygen [62]. Patients with respiratory distress will be made worse, organs and tissues of the whole body have different degrees of damage. Since the emergency epidemic, it is of high scientific significance to use bioinformatics to analyze the roles of novel coronavirus proteins. The study results show that some viral proteins could combine to the porphyrin to form a complex and at the same time, orf1ab, ORF3a, and ORF10 proteins could coordinate to attack the heme on hemoglobin. Lung cells are toxic and inflammatory derivatives produced by the attack, which eventually resulted in ground-glass-like lung damage [63]. Capillaries easily were broken due to inflammation. Proteins such as fibrinogen fill the capillaries' cracks through the coagulation reaction. Therefore, many fibrin accumulates in the alveolar tissue of the patient. As the porphyrin complexes of the virus produced in the human body, the mechanism also interfered with the normal heme anabolic pathway by a wide range of infection and disease. The virus may first infect cells with ACE2 receptors, including immune cells. Immune cells produced antibodies and viral proteins. Antibodies and red blood cells generated immune hemolysis, or red blood cells were infected by Spike-CD147 pathway [64]. The virus also captured porphyrin and inhibited heme metabolism and cytokine storm causes multiple organs failure. The protein, which acts as an ion channel, could be a target for new drugs against the SARS-CoV-2 virus [65].

The high expression of ACE2 in lateral ventricular choroid plexus further increases the possibility of SARS-CoV-2 invading the CNS [66]. Recently, the SARS-CoV-2 was also detected by genomic sequencing in CSF sample from a 24-year-old male COVID-19 patient in Japan [67]. The infection of SARS-CoV-2 in the CNS may produce unpredictable effect on neurodegenerative diseases. SARS-CoV-2 can activate glial cells, induce pro-inflammatory state, and even cause severe innate immune response and sustained increase in cytokine levels. In addition, persistent SARS-CoV-2 infection can also induce neuroimmune responses [68]. Retrospective studies have showed that corticosteroids were frequently used in the treatment of hospitalized COVID-19 patients [69], while it's worth noting that inappropri-

ate corticosteroid therapy produced adverse neuropsychiatric symptoms, affecting about 35% of COVID-19 patients, including cognitive and sleep disorders, delirium, hypomania, mania, depression, and psychosis [70]. Angiotensin-converting enzyme 2 is also highly expressed in dopaminergic neurons [71] and direct infection of SARS-CoV-2 upon neuronal cells, vast inflammatory agents induced by severe systemic inflammation flooding into brains leading to respiratory failure associated brain ischemia, thrombosis and stroke [72]. Nearly 30-60% of patients with COVID-19 suffer from CNS symptoms. Neuronal infection can be prevented by blocking ACE2 with antibodies or by administering cerebrospinal fluid from a COVID-19 patient [73]. In autopsies from patients who died of COVID-19, SARS-CoV-2 is detected in cortical neurons and pathological features associated with infection with minimal immune cell infiltrates [74].

The hypermetabolic state is unique to the SARS-CoV-2-infected cells [75] and highlights the ability of SARS-CoV-2 to hijack the host neuron machinery to replicate. It is confirmed that infection by SARS-CoV-2 induced a locally hypoxic environment in neuronal regions by staining for HIF1 α in mock-infected and SARS-CoV-2-infected organoids and it indicates the potential of SARS-CoV-2 in manipulating host metabolic programming, which may create a resource-restricted environment for the cells [76]. SARS-CoV2 infection have also developed a hyper inflammatory syndrome (also termed cytokine release syndrome), a dysfunction in autonomic tone to the cytokine release syndrome and related multiorgan damage in COVID-19 [77]. It is hypothesized that a cholinergic anti-inflammatory pathway could be targeted as a therapeutic avenue. This inflammatory response and the activated immune cells and inflammatory cytokines can spread through the bloodstream to cause multiorgan failure [78]. The cytokine release syndrome is likely an indirect effect of the virus and has been associated with rapid clinical deterioration leading to Acute Respiratory Distress Syndrome (ARDS) [79], multiorgan failure, and mortality. The COVID-19 syndrome results in the up-regulation of inflammasomes, dysregulation of T cells with associated lymphopenia, and unfettered production of cytokines/chemokines, including IL6, TNF α , and CCL2. The virus alters the microenvironment of the immune system, especially as it pertains to macrophages [80]. Highly inflammatory FCN1+ macrophages predominate over fatty acid-binding protein 4 (FABP4+) macrophages in patients with a more severe COVID-19 clinical course. In contrast, in milder cases of COVID-19, the expansion of clonal CD8+ T cells in the lung microenvironment suggests a robust adaptive immune response connected to a better control of COVID-19.

HLH (Hemophagocytic lymphohistiocytosis) is characterized by prolonged fever, cytopenia, and high serum ferritin, and is frequently associated with pulmonary disease [81]. Therefore, diagnosis and treatment of hyperinflammation have been suggested as a possible therapeutic approach. Consistent with this hypothesis, some experimental treatments (e.g., steroids, selective cytokine blockade with anakinra or tocilizumab) are currently being evaluated in clinical studies, despite their relative immunosuppressive effects. The pathological changes to various organ systems may be caused directly by the cytopathic effect mediated by SARS-CoV2 infecting cells expressing the angiotensin-converting enzyme 2 (ACE2) receptor. ACE2 counterbalances the function of ACE, which in return negatively regulates AngII production and may contribute to hyperinflammation [82,83]. ACE2 lowers blood pressure by catalyzing the hydrolysis of angiotensinII (vasoconstrictors) into angiotensin 1-9 and 1-7 respectively (vasodilators). Thus, a change in ACE2

function or level of expression can affect blood pressure and also neuronal regulation of inflammation and inhibition of the brain ACE2 function decreases the parasympathetic tone [84]. SARS-CoV infection has been associated with an impaired ACE2 expression and function. In mice, SARS-CoV infection considerably downregulates the ACE2 expression, including in the lungs [85,86]. The so-called ACE2 shedding process is tightly coupled with TNF α production. Notably, as part of a feedback loop, hyperactivation of the vagus nerve via the nicotinic acetylcholine receptor (nAChR) downregulates the expression or activity of ACE2 [87-89], which could prevent viral infection. Infections and danger-associated molecular patterns (DAMPs) can activate immune cells to produce inflammatory cytokines [90]. In turn, these inflammatory cytokines can activate the afferent sensory vagus nerve that transmits the information to the central nervous system where it is processed. If needed, the central nervous system can activate the efferent vagus nerve to control inflammation by inhibiting the production of inflammatory cytokines in macrophages [91]. Acetylcholine (ACh), the principal neurotransmitter of the vagus nerve, is well studied as a critical neurotransmitter but ACh also modulates immune cells. Acetylcholine is well conserved across evolution and is used by the nervous system to regulate organ function and physiological homeostasis including the immune system and inflammation. Indeed, it is reported that the vagus nerve, the longest nerve connecting the brain with most organs, controls immune cells by producing ACh [92]. This mechanism is so significant that some regulatory T lymphocytes replicate the nervous system through ACh synthesis in order to control cytokine production by macrophages. ACh can signal through either muscarinic or nicotinic receptors. The spleen is the target organ of the nicotinic-cholinergic system [93]. ACh inhibits the production of inflammatory cytokines in macrophages specifically via α 7 nicotinic ACh receptors (α 7nAChR) [94]. Specific inhibition of α 7nAChR in human macrophages causes overzealous production of inflammatory cytokines and prevents the potential of either ACh or nicotine to inhibit its production in human macrophages. Electrical stimulation of the vagus nerve in rodent models induces the production of ACh and inhibits systemic inflammation for wild-type but not α 7nAChR-KO mice with experimental sepsis induced by either endotoxemia or polymicrobial peritonitis [95-97]. Acute lung injury and ARDS are often derived from sepsis-induced pulmonary inflammation. Alveolar injury to the lungs as a result of mechanical ventilation (ventilator-induced lung injury) is enhanced by concomitant inflammation. Conversely, surgical vagotomy in animals undergoing mechanical ventilation increases the alveolar damage that is associated with robust production of inflammatory cytokines such as IL-6 [98]. Therefore, electrical or pharmacological vagal nerve activation attenuates lung injury by modulating the inflammatory response and subsequently cellular damage and organ injury and failure. Finally, Vagus Nerve Stimulation (VNS) also eases the downstream effects of cytokine activation on systemic coagulation in rodent models [99]. This supports the current prevailing concept that dysregulated endothelial inflammation upregulates thrombin generation in COVID-19-associated thrombus formation [100]. Patients with an increased vagal tone might be protected from a cytokine release syndrome [101]. The observation of lower rates of symptomatic COVID-19 infections in active smokers potentially suggests that active nicotine exposure activates the cholinergic anti-inflammatory pathway, previously shown to be protective in various infectious illnesses, despite the deleterious effects of tobaccos use. A milder COVID-19 disease course in children, who have a naturally higher vagal tone

[102], even in an infectious setting, could support the significance of the cholinergic anti-inflammatory pathway uniquely in COVID-19 patients.

Targeting the cholinergic anti-inflammatory pathway via vagus nerve stimulation

Importantly, nAChR plays a role in the expression of ACE2 [103,104] which has been identified as the key target receptor of SARS-CoV2 [105]. Thus, vagal stimulation can potentially prevent SARS-CoV2 propagation by inhibiting ACE2 expression. Vagal stimulation reduced blood C-Reactive Protein (CRP), fecal calprotectin, and abdominal pain, and improved mood. Vagal stimulation also improved clinical symptoms and significantly lowered serum levels of inflammatory cytokines such as TNF- α and IL-6 in 18 patients after 3 months of treatment. Clinically, VNS can be achieved pharmacologically or electrically via invasive cervical vagal stimulation or non-invasively via the ear or by electro acupuncture. An additional target for neuromodulation is the splenic nerve, which contains sympathetic and distal vagal parasympathetic nerve fibers to the spleen. In the experimental setting, the anti-inflammatory properties of the vagus nerve depend on the splenic nerve [106]. The splenic nerve stimulation exhibits comparable immunosuppressive effects as VNS and can be attempted percutaneously in an invasive and non-invasive fashion (ultrasound waves) [107]. To target the cholinergic anti-inflammatory pathway as a treatment for COVID-19 and the associated cytokine release syndrome, ongoing and future efforts will determine the potential utility of autonomic nerve modulation in the prevention and treatment of COVID-19 as shown in (Table 1) [108].

Table 1: Efforts to target the cholinergic anti-inflammatory pathway.

Strategy	Agent/technology
Pharmacological	Nicotine, GTS-21, a nAChR agonist
	Implantable cuffbased VNS
Neuromodulation	Transcutaneous non-invasive VNS via neck or ear
	Electroacupuncture
	Splenic nerve stimulation

VNS: vagus nerve stimulation; nAChR: nicotinic acetylcholine receptor.

In patients with COVID-19 and neurologic symptoms, an unusual pattern of marked CSF inflammation occurs, in which soluble markers were increased but white cell response and other immunologic features typical of CNS viral infections were absent [109]. These features distinguish COVID-19 CSF from other viral CNS infections and raise fundamental questions about the CNS pathobiology of SARS-CoV-2 infection. CSF biomarkers of intrathecal inflammation (CSF white blood cell count, neopterin, β 2-microglobulin, and immunoglobulin G index), blood-brain barrier integrity (albumin ratio), and axonal injury (CSF neurofilament light chain protein [NfL]) were assessed in 6 patients with moderate to severe coronavirus disease 2019 (COVID-19) and neurologic symptoms. CSF neopterin (median 43.0 nmol/L) and β 2-microglobulin (median 3.1 mg/L) were increased in all. Median immunoglobulin G index (0.39), albumin ratio (5.35), and CSF white blood cell count (<3 cells/ μ L) were normal, while CSF NfL was elevated in 2 patients. The ratio of albumin in Cerebrospinal Fluid (CSF) to serum may serve as an index of the integrity of the blood-CSF barrier, with increases in this ratio indicating increased permeability. The ratio of immunoglobulin G (IgG) in CSF to serum (divided by the albumin ratio to correct

for variance in blood-CSF permeability) represents an index of the endogenous production of IgG in the Central Nervous System (CNS), with increases reflecting a possible infectious and/or autoimmune process stimulating central IgG synthesis, CSF/serum albumin ratios or CSF/serum IgG indices [110].

Chemosensory Anatomy Defines the Potential Attack Surface for SARS-CoV-2. Chemosensation occurs in sensory epithelia in the nose and mouth. Multiple cranial nerves relay the senses of smell, taste, and chemesthesis to the brain. Airborne odors are detected by olfactory sensory neurons that reside in the olfactory epithelium; their axons pierce the bony cribriform plate to enter the olfactory bulb in the brain. Taste buds on the tongue are innervated by sensory afferents from the facial nerve (VII) and glossopharyngeal nerve (IX). The vagus nerve (X) also innervates taste buds residing in the pharynx. The detection of pungent chemicals, also known as chemesthesis, is mediated by both oral and nasal afferents of the trigeminal nerve (V). Although deficits in smell are most commonly reported in COVID-19, all three chemosensory modalities have been reported to be affected. Possible Mechanisms of Chemosensory Disturbances are [111].

- (A) COVID-19 patients have reported olfactory loss in the absence of features common to upper respiratory infections like widespread nasal inflammation or obstruction. Such symptoms are consistent with recent CT imaging suggesting that SARS-CoV-2 infection may cause inflammation that is localized to the olfactory clefts [112].
- (B) Sustentacular cells, Bowman’s gland cells, and microvillar cells in the olfactory epithelium express both ACE2 and TMPRSS2 and may be directly infected by SARS-CoV-2. Support cell infection may cause changes in the olfactory mucus or ion imbalances that can inhibit olfactory signaling; such changes may be rapidly reversible. The loss of these support cells in animal models can also result in the death of olfactory sensory neurons.
- (C) Inflammatory cytokines may also directly or indirectly inhibit olfactory sensory neuron function.
- (D) Although current data suggest that sustentacular and other support cells are the primary targets of SARS-CoV-2, a remaining possibility is that SARS-CoV-2 directly infects OSNs.
- (E) Immunostaining for ACE2 protein in the mouse olfactory bulb suggests that ACE2 expression is restricted to vascular pericytes [113], which may directly (via perfusion changes) or indirectly (via inflammation) affect chemosensory perception in the brain.
- (F) Taste and chemesthetic disturbances may result from the direct infection of cells in the tongue, the secondary consequences of obstruction due to inflammation, or damage following the release of inflammatory cytokines.

S proteins of SARS-CoV-2 and SARS-CoV are highly similar, sharing a sequence identity of 77% [114]. In addition, we found that the SARS-CoV-2 S protein is slightly more positively charged than that of SARS-CoV since it contains four more positively charged residues and five less negatively charged residues which may lead to an increased affinity to bind to negatively charged regions of other molecules through nonspecific and specific interactions. Analysis the S protein binding to the host ACE2 receptor showed a 30% higher binding energy for SARS-

CoV-2 than for the SARS-CoV S protein [115]. These results might be useful for understanding the mechanism of cell entry, blood-brain barrier crossing, and clinical features related to the CNS infection by SARS-CoV-2.

Brain and lung crosstalk during COVID-19 infection [116] as SAR-CoV-2 employs ACE2 as the receptor for viral cell entry and induction of lung injury through increasing the immune system cytokines. It can downregulate the central ACE2 protein expression; inhibition of ACE2 activity reduces the sensitivity of the baroreceptor reflex control of the heart rate as well as increases sympathetic tone which eventually results in the blood pressure elevation and cardiac dysfunction. In addition, concerning the neuroprotective property of ACE2, its downregulation may disturb the balance of neurotoxicity/neuroprotection inside the brain. Increase of inflammatory cytokines during lung injury, hypoxemia, and elevation of sympathetic tone through ACE2 downregulation leads to CNS hyperactivation which might play a crucial role in the etiopathogenesis of neurogenic pulmonary edema which may play a role in COVID-19 pulmonary complications [117] in patients.

COVID-19 initiates the inflammatory cascade and, as a result, releases inflammatory cytokines [118] which is called cytokine storm syndrome [119]. Consecutively, these cytokines can drive neuronal hyperexcitability via activation of glutamate receptors and play a role in the development of acute seizures [120-122]. Elevation of sympathetic tone through ACE2 downregulation leads to CNS hyperactivation which might play a crucial role in the etiopathogenesis of Neurogenic Pulmonary Edema (NPE) [123], a life-threatening complication following a neurologic insult [124], and finally leading to deterioration with the respiratory and cardiovascular complications in these patients.

The crucial step in the viral infection is the process of viral entry into the host cells. The endocytic pathway including endosomes and lysosomes and the autophagy process in viral entry has attracted. The presence of the virus in the brain stem may affect chemosensing neural cells related to respiration as well as respiratory center neurons, thus damaging the lung ventilatory function [125]. Spike protein and ACE2 represent the key, but not the exclusive, site of entry of the virus into the cell; thus, non-ACE2 pathways for virus infection of neural cells cannot be excluded. Whether COVID-19 infects neurons and astroglial cells and enters astrocytes by endocytosis remains to be studied. The Spike protein dependent pathway is thought to be more important than clathrin-dependent endocytosis for cell entry and BBB crossing. Therefore, the Spike dependent pathway should be taken into account in therapeutic strategies for specific antibodies or vaccine production research.

Research suggests that the virus may gain access to the brain via the forebrain's olfactory bulb, which is important for the processing of smell. Loss of smell is a symptom in many patients with COVID-19. The olfactory bulb is rich in the chemical dopamine, which is important for pleasure, motivation and action. It may be that COVID-19 alters the levels of dopamine [126] and other chemicals, such as serotonin and acetylcholine, in the brain. All these chemicals are known to be involved in attention, learning, memory and mood. These changes in the brain are likely responsible for the mood, fatigue and cognitive changes that are commonly experienced by COVID-19 patients [127]. This in turn may underlie the reported symptoms of stress, anxiety and depression in patients who have contracted the virus. But it's not just people who have contracted the COVID-19 virus that have suffered from increased anxiety and depression dur-

ing the pandemic. Excessive worry over contracting or spreading the virus to other family members, as well as isolation and loneliness, can also change our brain chemistry. Repeated stress is a major trigger for persistent inflammation in the body, which can also affect the brain and shrink the hippocampus and therefore affect the emotions. Stress can also affect levels of brain serotonin and cortisol, which can affect our mood. Eventually, these changes can cause symptoms of depression and anxiety [128]. The good thing about the brain, however, is that it is incredibly plastic, which means it is changeable and can compensate for damage. Memory loss and depression can be improved by doing things that alter the brain function and its chemistry [129].

Due to COVID-19 outbreak, few individuals were overwhelmed with pain & filled with great void because of being lonely and isolated at home. Aggression is a psychological construct in which the individual is not able to control mood impulses. It is closely linked to depression, suicidal tendencies, and substance abuse. In other words, such people show impulsive aggression towards oneself and others and become depressed under the stressful situations of life. Aggression & COVID-19 has emerged as a global public health crisis & threat. From the neurobiological point of view, there are structures, neuronal circuits that play a role [130]. To normalise the level of serotonin medical professionals can prescribe drugs against called as SSRs (Selective Serotonin Reuptake Inhibitors). Fluoxetine is most widely studied. β -adrenergic agonists are also one of the well-known drug used for dementia and brain injury. The neuronal center of aggression i.e. the Hypothalamus and the connections with the amygdala and hippocampus, located near the temporal lobe [131]. For the management of higher cognitive processes, frontal lobe plays an important role. i.e. regulations of emotions [132]. Centre for regulation of negative emotions, aggressive & violent behavior, impulsive acts, are situated in Prefrontal Cortex & the temporal regions of the brain. Experimentally induced anger in humans showed increased activation in the DLPFC (Dorsolateral Prefrontal Cortex) and OFC (Orbitofrontal Cortex). In non-human primates, OFC & DLPFC lesions reliably produce increased aggressive behavior. Studies also shows that Aggression in humans may be associated with decreased functioning in orbital and dorsolateral prefrontal cortical areas, and increased functioning in medial temporal (especially amygdala) brain regions. All neuronal pathways are complex and aggression in humans is due to decreased level of serotonin neurotransmitter. Sudden lockdown due to novel Corona virus (COVID-19) in rural area resulted in following consequences like lack of confidence, restlessness, lack of concentration, domestic violence- both verbal and non-verbal, nervousness, irritability, gender-based abuse, destructive thoughts, feeling to hit someone, frequent arguments at home, insecurity with spouse, violence at home and overthinking, suicidal cases & attempted suicide cases increased in lockdown period [133,134]. During quarantine period, counselling program is needed to prevent all these activities if people do not socialize, then the level of serotonin decreases leading to negative thoughts, depression, impulsive behavior, and arrogant nature. Even criminal tendency may develop in normal person for no reason. So, Serotonin plays a crucial role to maintain mood of person and it has been proved by genetic etiology. The polymorphic genetic variants of the serotonergic system are deeply affected by genetic predisposition that alters serotonin levels in the central & peripheral nervous system i.e., rate of serotonin production, synaptic release & pathways.

Chemical transmission between neurosecretory cells is

a central biological phenomenon. Detecting neurotransmitter molecules released in the brain or in cell cultures is key to understanding how, when, and where chemical transmission occurs. Carbon fiber microelectrodes provide a common basis for detecting not only dopamine but also a wide variety of neurotransmitters ranging from biogenic amines, purines, and amino acids to free radicals and peptides [135]. The neuro-immune pathway functions bidirectionally, where afferent neurons respond to immune signals at the periphery, and the efferent neurons promote the interaction between the brain and the periphery. This inflammatory reflex has been discussed viral invasion reaching the CNS through the olfactory bulb modulates brain cytokines in the brainstem, which could be related to elevation in the body temperature as well as disease progression [136]. The evidence of an exacerbated inflammatory response in severe cases of COVID-19 suggests that patients with olfactory and gustatory dysfunctions have less severe lung function impairments [137]. Viral genes involved in mechanisms such as biogenesis, entrance, replication, and infection are possible targets for host-cellular miRNAs and microRNAs (miRNAs) as possible therapeutic agents against SARS-CoV-2 [138]. The spike (S) protein of coronavirus, one of the surface glycoproteins, is divided into two functional units, S1 and S2. S1 facilitates virus infection by binding to host receptors, and S2 regulates the membranes fusion to enable viral RNA entering into host cells for further replication. Therefore, the S protein determines the host cell of the virus, regulates the viral attachment and fusion with the host cell membrane, and promotes cellular invasion. As such, the S protein is essential for viral infection, abnormal humoral metabolism is mainly manifested as imbalances of water and electrolytes [139]. Water and sodium disturbances along with the unusual serum potassium levels are most common. It has been shown that the incidence and severity of COVID-19 are closely related to abnormal metabolism of inorganic salts. Serum sodium shows a decrease trend in COVID-19 development. Hyponatremia (low blood sodium concentration) as well as low concentrations of potassium and calcium in the blood serum are also associated with COVID-19 [140]. Patients in Intensive Care Units (ICU) have higher plasma levels of IL2, IL7, IL10, MCP1, MIP1A, GSCF, IP10, and TNF α [141]. In these patients, white blood cells count, neutrophil count and D-dimer level keep rising while lymphocyte count keeps decreasing as the disease progresses [142]. Therefore, the infection along with the rapid replication of SARS-CoV-2 causes a large amount of body fluid permeating through pulmonary alveoli, leading to ARDS and lower expression of alveolar Na-K-ATPase promotes pulmonary edema [143].

Bradykinin (BK) is an important cellular mediator that causes vasodilatation and leaky blood vessels, leading to vascular leakage and edema. BK is generated by cleavage of High-Molecular-Weight Kininogen (HMWK) from plasma-kallikrein and binds to the BKB2 receptor, and thus results vascular hemorrhage. Inhibition of ACE2 by SARS-CoV-2 impairs the hydrolysis of des-arg9-bradykinin [144]. Therefore, the excessive release and decreased hydrolysis of BK through activating BKB1 and BKB2 receptors result in extra vascular leakage and pulmonary edema, inflammatory cytokines and inflammatory storm in COVID-19 patients can strongly induce the expression of HA-Synthase-2 (HAS2), while hyaluronidase level decreases, resulting in the accumulation of HA and inducing ARDS and pulmonary edema. The decreased expression of alveolar Na-K-ATPase, misregulation of sodium, potassium, AQP, and RAS channels and abnormal metabolism of BK and HA can all lead to lung liquid

clearance failure and pulmonary edema, resulting in severe lung damage and ARDS in COVID-19 patients [145].

Immune-modulatory agents for COVID-19 include tocilizumab, human immunoglobulin and the convalescent plasma. IL6 monoclonal antibody or tocilizumab was thought to work by interrupting inflammatory storm after the infection, but the latest clinical study published in NEJM showed that Tocilizumab was not effective in preventing intubation or death in mild hospitalized COVID-19 patients. Convalescent plasma have been initially shown to be beneficial for COVID-19 patients with severe infection stabilizing the immune system, but the subsequent randomized controlled trial did not show significant improvement within 28 days [146].

Targeting sodium channels and pumps

Since the inhibition of ENaC induces pulmonary edema formation, targeting ENaC is rational in order to enhance fluid clearance from the alveoli. Studies showed that ENaC activators or stimulators can regulate ENaC-dependent fluid absorption in alveolar and pulmonary edema [147]. Activation of β -adrenergic receptor, especially β 2, was found to stimulate Na⁺ and fluid reabsorption. It was observed that the expression of ENaC and Na⁺/K⁺-ATPase in primary alveolar type II cells from rat lungs increased responding to terbutaline. Inhalation or infusion of salbutamol, a β 2-adrenergic agonist, reduced the incidences of pulmonary edema and was found to be beneficial in ARDS patients [148]. Amiloride, a prototypic inhibitor of ENaC, might also have a potential in treating COVID-19 patients. Amiloride was shown to induce the reduction of ACE2 expression in bronchial and alveolar epithelial cells and can counteract the low cytosolic pH which has been observed in COVID-19 patients by acting on Na⁺/H⁺ exchanger [149]. Glucocorticoids can reduce pulmonary edema by regulating fluid absorption in alveoli and can be used as anti-inflammatory drugs in ARDS and pulmonary edema as well as in COVID-19 patients [150].

CFTR (Cystic Fibrosis Transmembrane Conductance Regulator) and NKCC (Na-K-2Cl cotransporter) inhibitors show promise in treating pulmonary edema. Furosemide, a NKCC inhibitor, has been acknowledged as first-line therapeutic drug for pulmonary edema in all the time [151]. CFTR inhibitors, like glibenclamide and CFTRinh-172 inhibitor, distinctly reduced absorptive alveolar fluid transport [152,153].

References

1. Du L, He Y, Zhou Y, Liu S, Zheng B J, et al. The spike protein of SARS-CoV-a target for vaccine and therapeutic development. *Nature Reviews Microbiology*. 2009; 7: 226-236
2. Perlman S, Netland J. Coronaviruses post-SARS: Update on replication and pathogenesis. *Nature Reviews Microbiology*. 2009; 7: 439-450.
3. Cooper KW, Brann DH, Farruggia MC, Bhutani S, Pellegrino R, et al. COVID-19 and the Chemical Senses: Supporting Players Take Center Stage. *Neuron*. 2020; 107(2): 219-233.
4. Pellegrino R, Cooper KW, Di Pizio A, Joseph PV, Bhutani S, et al. Corona Viruses and the Chemical Senses: Past, Present, and Future. *Chemical Senses*. 2020.
5. Wood Alex, Linley P, Maltby John, Kashdan Todd, Hurling Robert. Using personal and psychological strengths leads to increases in well-being over time: A longitudinal study and the development of the Strengths Use Questionnaire. *Personality and Individual*

- Differences. 2010; 50: 1.
6. Hu T, Liu Y, Zhao M, Zhuang Q, Xu L, et al. A comparison of COVID-19, SARS and MERS. *Peer Journal*. 2020; 19(8): e9725.
 7. Hwang WC. The psychotherapy adaptation and modification framework: Application to Asian Americans. *American Psychologist*. 2006; 61(7): 702-715.
 8. Ke Z, Oton J, Qu K, et al. Structures and distributions of SARS-CoV-2 spike proteins on intact virions. *Nature*. 2020; 588: 498-502.
 9. Wang MY, Zhao R, Gao LJ, Gao XF, Wang DP, et al. SARS-CoV-2: Structure, Biology, and Structure-Based Therapeutics Development. *Frontiers in Cellular And Infection Microbiology*. 2020; 10: 587269.
 10. Mulay A, Konda B, Garcia G Jr, Yao C, Beil S, et al. SARS-CoV-2 infection of primary human lung epithelium for COVID-19 modeling and drug discovery. *Cell Reports*. 2021; 35(5): 109055.
 11. Markus Hoffmann, Hannah Kleine-Weber, Simon Schroeder, Nadine Krüger, Tanja Herrler, et al. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell*. 2020; 181(2): 271-280.
 12. Von Bartheld, Christopher Hagen, Molly Butowt, Rafal. (2020). Prevalence of Chemosensory Dysfunction in COVID-19 Patients: A Systematic Review and Meta-analysis Reveals Significant Ethnic Differences. *medRxiv: The preprint server for health sciences*. 2020. 10.1101/2020.06.15.20132134.
 13. Minh P, Hoang Phillip, Staibano Tobial, Mc Hugh, Doron D, et al. Self-reported olfactory and gustatory dysfunction and psychophysical testing in screening for COVID-19: A systematic review and meta-analysis, *International Forum of Allergy & Rhinology*. 2021; 12(5): 744-756.
 14. Liang S W, Zhao J B, Zhao J B. Effect of mental-health curriculum on the emotion and attitudes toward seeking professional help of college students. *Chinese Journal of Health Education*. 2017; 33: 745-748.
 15. Butowt R, Von Bartheld CS. Anosmia in COVID-19: Underlying Mechanisms and Assessment of an Olfactory Route to Brain Infection. *Neuroscientist*. 2021; 27(6): 582-603.
 16. Login H, Håglin S, Berghard A, Bohm S. The Stimulus-Dependent Gradient of Cyp26B1+ Olfactory Sensory Neurons Is Necessary for the Functional Integrity of the Olfactory Sensory Map. *Journal of Neuroscience*. 2015; 35(40): 13807-13818
 17. Meinhardt J, Radke J, Dittmayer C, et al. Olfactory transmucosal SARS-CoV-2 invasion as a port of central nervous system entry in individuals with COVID-19. *Nature Neuroscience*. 2021; 24: 168-175.
 18. Bryche B, St Albin A, Murri S, Lacôte S, Pulido C, et al. Massive transient damage of the olfactory epithelium associated with infection of sustentacular cells by SARS-CoV-2 in golden Syrian hamsters. *Brain, Behaviour, And Immunity*. 2020; 89: 579-586.
 19. Sia SF, Yan LM, Chin AWH, et al. Pathogenesis and transmission of SARS-CoV-2 in golden hamsters. *Nature*. 2020; 583: 834-838
 20. Ueha R, Ito T, Furukawa R, Kitabatake M, Oujii-Sageshima N, et al. Oral SARS-CoV-2 Inoculation Causes Nasal Viral Infection Leading to Olfactory Bulb Infection: An Experimental Study. *Frontiers in Cellular and Infection Microbiology*. 2022; 12: 924725.
 21. Li Yan-Chao, Bai Wan Zhu, Hirano Norio, Hayashida Tsuyako, Taniguchi Takahide, et al. Neurotropic virus tracing suggests a membranous-coating-mediated mechanism for transsynaptic communication. *The Journal of comparative neurology*. 2013; 521(1): 203-212
 22. Eliezer Michael, Hautefort Charlotte, Hamel Anne-Laure, Verilaud Benjamin, Herman Philippe, et al. Sudden and Complete Olfactory Loss Function as a Possible Symptom of COVID-19. *JAMA Otolaryngology - Head and Neck Surgery*. 2020; 146.
 23. Chen ZS, Wong AKY, Cheng TC, Koon AC, Chan HYE. FipoQ/FBXO33, a Cullin-1-based ubiquitin ligase complex component modulates ubiquitination and solubility of polyglutamine disease protein. *Journal of Neurochemistry*. 2019; 149(6): 781-798.
 24. Torabi A, Mohammadbagheri E, Akbari Dilmaghani N, Bayat A H, Fathi M, et al. Proinflammatory Cytokines in the Olfactory Mucosa Result in COVID-19 Induced Anosmia. *ACS Chemical Neuroscience*. 2020; 11(13): 1909-1913.
 25. Rodríguez-Rey R, Garrido-Hernansaiz H, Collado S. Psychological Impact and Associated Factors during the Initial Stage of the Coronavirus (COVID-19) Pandemic among the General Population in Spain. *Frontiers in Psychology*. 2020; 11: 1540
 26. He X, Lau EHY, Wu P, et al. Temporal dynamics in viral shedding and transmissibility of COVID-19. *Nature Medicine*. 2020; 26: 672-675.
 27. Annika Armulik, Guillem Genové, Christer Betsholtz. Pericytes: Developmental, Physiological, and Pathological Perspectives, Problems, and Promises, *Developmental Cell*. 2011; 21(2): 193-215.
 28. Han Qi, Peng Jiakuan, Xu Hao, Chen Qianming. Taste Cell Is Abundant in the Expression of ACE2 Receptor of 2019-nCoV. 2020. 10.20944/preprints202004.0424.v1.
 29. Barlow LA. Progress and renewal in gustation: New insights into taste bud development. *Developmentm*. 2015; 142(21): 3620-3629.
 30. Frasnelli Johannes, Manescu Simona. The Intranasal Trigeminal System. 2017. 10.1007/978-3-319-26932-0_46.
 31. Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, et al. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science*. 2020; 370(6518): 856-860.
 32. Van Kampen JJA, Van De Vijver, DAMC, Fraaij PLA, et al. Duration and key determinants of infectious virus shedding in hospitalized patients with coronavirus disease-2019 (COVID-19). *Nature Communications*. 2021; 12: 267.
 33. Verdecchia P, Cavallini C, Spanevello A, Angeli F. The pivotal link between ACE2 deficiency and SARS-CoV-2 infection. *European Journal of Internal Medicine*. 2020; 76: 14-20.
 34. Muus C, Luecken MD, Eraslan G, et al. Single-cell meta-analysis of SARS-CoV-2 entry genes across tissues and demographics. *Nature Medicine*. 2021; 27: 546-559
 35. González-Duarte A, Norcliffe-Kaufmann L. Is happy hypoxia in COVID-19 a disorder of autonomic interoception? A hypothesis. *Clinical Autonomic Research*. 2020; 30(4): 331-333
 36. Hasanoglu I, Korukluoglu G, Asilturk D, Cosgun Y, Kalem AK, et al. Higher viral loads in asymptomatic COVID-19 patients might be the invisible part of the iceberg. *Infection*. 2021; 49(1): 117-126.
 37. Puelles VG, Lütgehetmann M, Lindenmeyer MT, Sperhake JP, Wong MN, et al. Multiorgan and Renal Tropism of SARS-CoV-2. *New England Journal of Medicine*. 2020; 383(6): 590-592.
 38. Chen N, Zhou M, Dong X, Qu J, Gong F, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: A descriptive study. *Lancet*. 2020; 395(10223): 507-513.

39. Ciaccio M, Agnello L. Biochemical biomarkers alterations in Coronavirus Disease 2019 (COVID-19), Diagnosis (Berlin). 2020; 7(4): 365-372.
40. Yadav R, Chaudhary JK, Jain N, Chaudhary PK, Khanra S, et al. Role of Structural and Non-Structural Proteins and Therapeutic Targets of SARS-CoV-2 for COVID-19. *Cells*. 2021; 10(4): 821.
41. Saksena N, Bonam SR, Miranda-Saksena M. Epigenetic Lens to Visualize the Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2) Infection in COVID-19 Pandemic. *Frontiers in Genetics*. 2021; 12: 581726.
42. Zhou Z, Kang H, Li S, Zhao X. Understanding the neurotropic characteristics of SARS-CoV-2: from neurological manifestations of COVID-19 to potential neurotropic mechanisms. *Journal of Neurology*. 2020; 267(8): 2179-2184.
43. Montaldo C, Messina F, Abbate I, Antonioli M, Bordoni V, et al. Multi-omics approach to COVID-19: a domain-based literature review. *Journal of Translational Medicine*. 2021; 19(1): 501
44. Zubair AS, McAlpine LS, Gardin T, Farhadian S, Kuruvilla DE, et al. Neuropathogenesis and Neurologic Manifestations of the Coronavirus in the Age of Coronavirus Disease 2019: A Review. *JAMA Neurology*. 2020; 77(8): 1018-1027.
45. Pennisi M, Lanza G, Falzone L, Fiscaro F, Ferri R, et al. SARS-CoV-2 and the Nervous System: From Clinical Features to Molecular Mechanisms. *International Journal of Molecular Sciences*. 2020; 21(15): 5475.
46. Li YX, Bai WZ, Hashikawa T. The neuroinvasive potential of SARS-CoV2 may be at least partially responsible for the respiratory failure of COVID-19 patients. *Journal of Medical Virology*. 2020.
47. Guillaud L, El-Agamy SE, Otsuki M, Terenzio M. Anterograde Axonal Transport in Neuronal Homeostasis and Disease. *Frontiers in Molecular Neuroscience*. 2020; 13: 556175.
48. Severo bem junior, Do rego aquino P, Nunes rabelo N, Do rego aquino M, Veiga silva A, et al. Sars-cov-2 and Nervous System - Neurological Manifestations in Patients with COVID-19: A Systematic Review. *Journal of Neurology Research, North America*. 2020; 10(4): 113-121
49. Shen XR, Geng R, Li Q, Chen Y, Li SF, et al. ACE2-independent infection of T lymphocytes by SARS-CoV-2. *Signal Transduction And Targeted Therapy*. 2022; 7(1): 83.
50. Frontera JA, Melmed K, Fang T, et al. Toxic Metabolic Encephalopathy in Hospitalized Patients with COVID-19. *Neurocritical Care*. 2021; 35: 693-706.
51. Kanberg N, Ashton NJ, Andersson LM, Yilmaz A, Lindh M, et al. Neurochemical evidence of astrocytic and neuronal injury commonly found in COVID-19. *Neurology*. 2020; 95(12): e1754-e1759.
52. Virhammar J, Nääs A, Fällmar D, Cunningham JL, Klang A, et al. Biomarkers for central nervous system injury in cerebrospinal fluid are elevated in COVID-19 and associated with neurological symptoms and disease severity. *European Journal of Neurology*. 2021; 28(10): 3324-3331.
53. Espindola Otavio, Brandão Carlos, Gomes Yago, Gomes Pinheiro, Siqueira Marilda, et al. Cerebrospinal fluid findings in neurological diseases associated with COVID-19 and insights into mechanisms of disease development. *International Journal of Infectious Diseases*. 2021; 102: 155-162
54. Singh AK, Singh A, Shaikh A, Singh R, Misra A. Chloroquine and hydroxychloroquine in the treatment of COVID-19 with or without diabetes: A systematic search and a narrative review with a special reference to India and other developing countries. *Diabetes & Metabolic Syndrome*. 2020; 14(3): 241-246.
55. Saghir SAM, AlGabri NA, Alagawany MM, Attia YA, Alyileili SR, et al. Chloroquine and Hydroxychloroquine for the Prevention and Treatment of COVID-19: A Fiction, Hope or Hype? An Updated Review. *Therapeutics and Clinical Risk Management*. 2021; 17: 371-387.
56. Jain S, Potschka H, Chandra PP, Tripathi M, Vohora D. Management of COVID-19 in patients with seizures: Mechanisms of action of potential COVID-19 drug treatments and consideration for potential drug-drug interactions with anti-seizure medications. *Epilepsy Research*. 2021; 174: 106675.
57. Omolo CA, Soni N, Fasiku VO, Mackraj I, Govender T. Update on therapeutic approaches and emerging therapies for SARS-CoV-2 virus. *European Journal of Pharmacology*. 2020; 883: 173348.
58. Pandey A, Nikam AN, Shreya AB, Mutalik SP, Gopalan D, et al. Potential therapeutic targets for combating SARS-CoV-2: Drug repurposing, clinical trials and recent advancements. *Life Sciences*. 2020; 256: 117883
59. Saha, Abinit, Sharma, Ashish, Bhattacharya, et al. Probable Molecular Mechanism of Remdesivir for the Treatment of COVID-19: Need to Know More. *Archives of Medical Research*. 2020; 51. 10.1016/j.arcmed.2020.05.001.
60. Wenzhong liu, hualan L. COVID-19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism. *ChemRxiv*. Cambridge: Cambridge Open Engage, Biological and Medicinal Chemistry. 2020.
61. Casqueiro J, Casqueiro J, Alves C. Infections in patients with diabetes mellitus: A review of pathogenesis. *Indian Journal of Endocrinology and Metabolism*. 2012; Suppl 1(Suppl1): S27-36
62. Clementi N, Ghosh S, De Santis M, Castelli M, Crisculo E, et al. Viral Respiratory Pathogens and Lung Injury. *Clinical Microbiology Reviews*. 2021; 34(3): e00103-20
63. Kronstein-Wiedemann R, Stadtmüller M, Traikov S, Georgi M, Teichert M, et al. SARS-CoV-2 Infects Red Blood Cell Progenitors and Dysregulates Hemoglobin and Iron Metabolism. *Stem Cell Reviews and Reports*. 2022; 18(5): 1809-1821.
64. Singh Tomar PP, Arkin IT. SARS-CoV-2 E protein is a potential ion channel that can be inhibited by Gliclazide and Memantine. *Biochemical and Biophysical Research Communications*. 2020; 530(1): 10-14.
65. Hu C, Chen C, Dong XP. Impact of COVID-19 Pandemic on Patients with Neurodegenerative Diseases. *Frontiers in Aging Neuroscience*. 2021; 13: 664965.
66. Moriguchi T, Harii N, Goto J, Harada D, Sugawara H, et al. A first case of meningitis/encephalitis associated with SARS-Coronavirus-2. *International Journal of Infectious Diseases*. 2020; 94: 55-58.
67. Beyerstedt S, Casaro EB, Rangel ÉB. COVID-19: angiotensin-converting enzyme 2 (ACE2) expression and tissue susceptibility to SARS-CoV-2 infection. *European Journal of Clinical Microbiology & Infectious Diseases*. 2021; 40(5): 905-919
68. Ur A, Verma K. Cytokine Storm in COVID19: A Neural Hypothesis. *ACS Chemical Neuroscience*. 2020; 11: 1868-1870
69. Van Paassen J, Vos JS, Hoekstra EM, et al. Corticosteroid use in COVID-19 patients: A systematic review and meta-analysis on clinical outcomes. *Critical Care*. 2020; 24: 696
70. García CAC, Sánchez EBA, Huerta DH, Gómez-Arnau J. Covid-19 treatment-induced neuropsychiatric adverse effects. *General Hospital Psychiatry*. 2020; 67: 163-164.

71. Valenzuela R, Rodriguez-Perez AI, Costa-Besada MA, Rivas-Santesteban R, Garrido-Gil P, et al. An ACE2/Mas-related receptor MrgE axis in dopaminergic neuron mitochondria. *Redox Biology*. 2021; 102078.
72. Szelenger R, Saluk-Bijak J, Bijak M. Ischemic Stroke among the Symptoms Caused by the COVID-19 Infection. *Journal of Clinical Medicine*. 2020; 9(9): 2688.
73. Song E, Zhang C, Israelow B, Lu-Culligan A, Prado AV, et al. Neuroinvasion of SARS-CoV-2 in human and mouse brain. *Journal of Experimental Medicine*. 2021; 218(3): e20202135.
74. Satturwar S, Fowkes M, Farver C, Wilson AM, Eccher A, et al. Postmortem Findings Associated with SARS-CoV-2: Systematic Review and Meta-analysis. *American Journal of Surgical Pathology*. 2021; 45(5): 587-603.
75. Seehusen F, Clark JJ, Sharma P, Bentley EG, Kirby A, et al. Neuroinvasion and Neurotropism by SARS-CoV-2 Variants in the K18-hACE2 Mouse. *Viruses*. 2022; 14: 1020.
76. Serebrovska ZO, Chong EY, Serebrovska TV, et al. Hypoxia, HIF-1 α , and COVID-19: from pathogenic factors to potential therapeutic targets. *Acta Pharmacologica Sinica*. 2020; 41: 1539-1546
77. Chigr F, Merzouki M, Najimi M. Autonomic brain centers and pathophysiology of COVID-19. *ACS Chemical Neuroscience*. 2020; 11: 1520-1522.
78. Fudim M, Qadri YJ, Ghadimi K, MacLeod DB, Molinger J, et al. Implications for Neuromodulation Therapy to Control Inflammation and Related Organ Dysfunction in COVID-19. *Journal of Cardiovascular Translational Research*. 2020; 13(6): 894-899.
79. Hojyo S, Uchida M, Tanaka K, et al. How COVID-19 induces cytokine storm with high mortality. *Inflammation and Regeneration*. 2020; 40: 37.
80. Liao M, Liu Y, Yuan J, et al. The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA sequencing. *medRxiv*. 2020.
81. Mehta P, McAuley D F, Brown M, et al. COVID-19: Consider cytokine storm syndromes and immunosuppression. *Lancet*. 2020; 395: 1033-1034.
82. Potter E K. Angiotensin inhibits action of vagus nerve at the heart. *British Journal of Pharmacology*. 1982; 75: 9-11
83. Xu Z, Li W, Han J, et al. Angiotensin II induces kidney inflammatory injury and fibrosis through binding to myeloid differentiation protein-2 (MD2). *Scientific Reports*. 2017; 7: 44911.
84. Xia H, Feng Y, Obr T D, Hickman P J, Lazartigues E. Angiotensin II type 1 receptor-mediated reduction of angiotensin-converting enzyme 2 activity in the brain impairs baroreflex function in hypertensive mice. *Hypertension*. 2009; 53: 210-216.
85. Haga S, Yamamoto N, Nakai-Murakami C, et al. Modulation of TNF-alpha-converting enzyme by the spike protein of SARS-CoV and ACE2 induces TNF-alpha production and facilitates viral entry. *Proceedings of the National Academy of Sciences of the United States of America*. 2008; 105: 7809-7814.
86. Kuba K, Imai Y, Rao S, et al. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nature Medicine*. 2005; 11: 875-879.
87. Oakes J M, Fuchs R M, Gardner J D, Lazartigues E, Yue X. Nicotine and the renin-angiotensin system. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*. 2018; 315: R895-R906.
88. Ferrari M F, Raizada M K, Fior-Chadi DR. Differential regulation of the renin-angiotensin system by nicotine in WKY and SHR glia. *Journal of Molecular Neuroscience*. 2008; 35: 151-160.
89. Ferrari M F, Raizada M K, Fior-Chadi D R. Nicotine modulates the renin-angiotensin system of cultured neurons and glial cells from cardiovascular brain areas of Wistar Kyoto and spontaneously hypertensive rats. *Journal of Molecular Neuroscience*. 2007; 33: 284-293.
90. Lotze M T, Tracey K J. High-mobility group box 1 protein (HMGB1): Nuclear weapon in the immune arsenal. *Nature Reviews. Immunology*. 2005; 5: 331-342.
91. Tracey K J. The inflammatory reflex. *Nature*. 2002; 420: 853-859.
92. Bassi G S, Kanashiro A, Coimbra N C, Terrando N, Maixner W, et al. Anatomical and clinical implications of vagal modulation of the spleen. *Neuroscience and Biobehavioral Reviews*. 2020; 112: 363-373.
93. Huston J M, Ochani M, Rosas-Ballina M, et al. Splenectomy inactivates the cholinergic antiinflammatory pathway during lethal endotoxemia and polymicrobial sepsis. *The Journal of Experimental Medicine*. 2006; 203: 1623-1628.
94. Ulloa L. The vagus nerve and the nicotinic anti-inflammatory pathway. *Nature Reviews. Drug Discovery*. 2005; 4: 673-684.
95. Li W, Olshansky B. Inflammatory cytokines and nitric oxide in heart failure and potential modulation by vagus nerve stimulation. *Heart Failure Reviews*. 2011; 16: 137-145.
96. Zhao Y X, He W, Jing X H, et al. Transcutaneous auricular vagus nerve stimulation protects endotoxemic rat from lipopolysaccharide-induced inflammation. *Evidence-based Complementary and Alternative Medicine*. 2012; 627023.
97. Song X M, Li J G, Wang Y L, et al. The protective effect of the cholinergic anti-inflammatory pathway against septic shock in rats. *Shock*. 2008; 30: 468-472.
98. Dos Santos C C, Shan Y, Akram A, Slutsky A S, Haitsma J J. Neuro-immune regulation of ventilator-induced lung injury. *American Journal of Respiratory and Critical Care Medicine*. 2011; 183: 471-482.
99. Van Westerloo D J, Giebelen I A, Meijers J C, et al. Vagus nerve stimulation inhibits activation of coagulation and fibrinolysis during endotoxemia in rats. *Journal of Thrombosis and Haemostasis*. 2006; 4: 1997-2002.
100. Connors J M, Levy J H. Thromboinflammation and the hypercoagulability of COVID-19. *Journal of Thrombosis and Haemostasis*. 2020.
101. Huston J M, Tracey K J. The pulse of inflammation: heart rate variability, the cholinergic anti-inflammatory pathway and implications for therapy. *Journal of Internal Medicine*. 2011; 269: 45-53
102. Estevez-Baez M, Carricarte-Naranjo C, Jas-Garcia J D, et al. Influence of heart rate, age, and gender on heart rate variability in adolescents and young adults. *Advances in Experimental Medicine and Biology*. 2019; 1133: 19-33.
103. Oakes JM, Fuchs RM, Gardner JD, Lazartigues E, Yue X. Nicotine and the renin-angiotensin system. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*. 2018; 315: R895-R906.
104. Ferrari MF, Raizada MK, Fior-Chadi DR. Differential regulation of the renin-angiotensin system by nicotine in WKY and SHR glia. *Journal of Molecular Neuroscience*. 2008; 35: 151-160.
105. Hoffmann M, Kleine-Weber H, Schroeder S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020; 181: 271-280.

106. Vida G, Pena G, Deitch EA, Ulloa L. Alpha7-cholinergic receptor mediates vagal induction of splenic norepinephrine. *Journal of Immunology*. 2011; 186: 4340-4346.
107. Zachs DP, Offutt SJ, Graham RS, et al. Noninvasive ultrasound stimulation of the spleen to treat inflammatory arthritis. *Nature Communications*. 2019; 10: 951.
108. Fudim M, Qadri YJ, Ghadimi K, MacLeod DB, Molinger J, et al. Implications for Neuromodulation Therapy to Control Inflammation and Related Organ Dysfunction in COVID-19. *Journal of Cardiovascular Translational Research*. 2020; 13(6): 894-899.
109. Edén A, Kanberg N, Gostner J, Fuchs D, Hagberg L, et al. CSF Biomarkers in Patients With COVID-19 and Neurologic Symptoms: A Case Series. *Neurology*. 2021; 96(2): e294-e300.
110. Lewis A, Frontera J, Placantonakis DG, Lighter J, Galetta S, et al. Cerebrospinal fluid in COVID-19: A systematic review of the literature. *Journal of Neurological Sciences*. 2021; 421: 117316.
111. Cooper KW, Brann DH, Farruggia MC, Bhutani S, Pellegrino R, et al. COVID-19 and the Chemical Senses: Supporting Players Take Center Stage. *Neuron*. 2020; 107(2): 219-233.
112. Eliezer M, Hautefort C, Hamel AL, Verillaud B, Herman P, et al. Sudden and Complete Olfactory Loss Function as a Possible Symptom of COVID-19. *JAMA Otolaryngology. Head Neck Surgery*. 2020.
113. Brann D, Tsukahara T, Weinreb C, Lipovsek M, Van den Berge K, et al. Non-neuronal expression of SARS-CoV-2 entry genes in the olfactory system suggests mechanisms underlying COVID-19-associated anosmia. *bioRxiv*. 2020.
114. Hassanzadeh K, Perez Pena H, Dragotto J, Buccarello L, Iorio F, et al. Considerations around the SARS-CoV-2 Spike Protein with Particular Attention to COVID-19 Brain Infection and Neurological Symptoms. *ACS Chemical Neuroscience*. 2020; 11(15): 2361-2369.
115. Ortega JT, Serrano ML, Pujol FH, Rangel HR. Role of changes in SARS-CoV-2 spike protein in the interaction with the human ACE2 receptor: An in silico analysis. *EXCLI Journal*. 2020; 19: 410-417.
116. Robba C, Battaglini D, Ball L, Patroniti N, Loconte M, et al. Distinct phenotypes require distinct respiratory management strategies in severe COVID-19. *Respiratory Physiology & Neurobiology*. 2020; 279: 03455.
117. Chen LD, Zhang ZY, Wei XJ, et al. Association between cytokine profiles and lung injury in COVID-19 pneumonia. *Respir Res*. 2020; 21: 201.
118. Huang C, Wang Y, Li X, Ren L, Zhao J, et al. Clinical Features of Patients Infected with 2019 Novel Coronavirus in Wuhan, China. *Lancet*. 2020; 395(10223): 497-506
119. Zhao M. Cytokine Storm and Immunomodulatory Therapy in COVID-19: Role of Chloroquine and Anti-IL-6 Monoclonal Antibodies. *International Journal of Antimicrobial Agents*. 2020; 55: 105982.
120. Singhi P. Infectious Causes of Seizures and Epilepsy in the Developing World. *Developmental Medicine & Child Neurology*. 2011; 53(7): 600-609.
121. Libbey J E, Kennett N J, Wilcox K S, White H S, Fujinami R S. Interleukin-6, Produced by Resident Cells of the Central Nervous System and Infiltrating Cells, Contributes to the Development of Seizures Following Viral Infection. *Journal of Virology*. 2011; 85(14): 6913-6922.
122. Libbey J E, Kirkman N J, Smith M C P, Tanaka T, Wilcox K S, et al. Seizures Following Picornavirus Infection. *Epilepsia*. 2008; 49(6): 1066-1074.
123. Terrence C F, Rao G R, Perper JA. Neurogenic Pulmonary Edema in Unexpected, Unexplained Death of Epileptic Patients. *Annals of Neurology*. 1981; 9(5): 458-464.
124. Šedý J, Kunes J, Zicha J. Pathogenetic Mechanisms of Neurogenic Pulmonary Edema. *Journal of Neurotrauma*. 2015; 32(15): 1135-1145.
125. Steardo L, Steardo L, Zorec R, Verkhatsky A. Neuroinfection May Contribute to Pathophysiology and Clinical Manifestations of COVID-19. *Acta Physiologica*. 2020; 229: e13473.
126. Nataf S. An alteration of the dopamine synthetic pathway is possibly involved in the pathophysiology of COVID-19. *Journal of Medical Virology*. 2020.
127. Rogers JP, Chesney E, Oliver D, Pollak TA, McGuire P, et al. Psychiatric and neuropsychiatric presentations associated with severe coronavirus infections: a systematic review and meta-analysis with comparison to the COVID-19 pandemic. *Lancet Psychiatry*. 2020; 7(7): 611-627.
128. Adhanom Ghebreyesus T. Addressing mental health needs: An integral part of COVID-19 response. *World Psychiatry*. 2020; 19(2): 129-130
129. Boldrini M, Canoll PD, Klein RS. How COVID-19 Affects the Brain. *JAMA Psychiatry*. 2021; 78(6): 682-683
130. Mazza M, Marano G, Lai C, Janiri L, Sani G. Danger in danger: Interpersonal violence during COVID-19 quarantine. *Psychiatry Research*. 2020; 289: 113046.
131. Šimić G, Tkalčić M, Vukić V, Mulc D, Španić E, et al. Understanding Emotions: Origins and Roles of the Amygdala. *Biomolecules*. 2021; 11(6): 823.
132. Hoaken, Peter, Allaby, David, Earle, et al. Executive cognitive functioning and the recognition of facial expressions of emotion in incarcerated violent offenders, non-violent offenders, and controls. *Aggressive behavior*. 2007; 33: 412-21
133. Sharma A, Borah SB. Covid-19 and Domestic Violence: An Indirect Path to Social and Economic Crisis. *Journal of Family Violence*. 2022; 37(5): 759-765.
134. Jesus, Tiago Bhattacharjya, Sutanuka, Papadimitriou, Christina, et al. Lockdown-Related Disparities Experienced by People with Disabilities during the First Wave of the COVID-19 Pandemic: Scoping Review with Thematic Analysis. *International Journal of Environmental Research and Public Health*. 2021; 18: 6178.
135. Huffman ML, Venton BJ. Carbon-fiber microelectrodes for in vivo applications. *Analyst*. 2009; 134(1): 18-24.
136. Pavlov VA, Tracey KJ. The vagus nerve and the inflammatory reflex—linking immunity and metabolism. *Nature Reviews Endocrinology*. 2012; 8(12): 743-54.
137. Gupta V, Banavara Rajanna L, Upadhyay K, Bhatia R, Madhav Reddy N, Malik D, Srivastava A. Olfactory and Gustatory Dysfunction in COVID-19 Patients from Northern India: A Cross-Sectional Observational Study. *Indian Journal of Otolaryngology and Head & Neck Surgery*. 2021; 73(2): 218-225
138. Ahmed JQ, Maulud SQ, Dhawan M, Priyanka Choudhary, OP Jalal, et al. Micro RNAs in the development of potential therapeutic targets against COVID-19: A narrative review. *Journal of Infection and Public Health*. 2022; 15(7): 788-799.
139. Lippi G, South AM, Henry BM. Electrolyte imbalances in patients with severe coronavirus disease 2019 (COVID-19). *Annals of Clinical Biochemistry*. 2020; 57(3): 262-26.

140. Hu W, Lv X, Li C, Xu Y, Qi Y, et al. Disorders of sodium balance and its clinical implications in COVID-19 patients: A multicenter retrospective study. *Internal and Emergency Medicine*. 2021; 16(4): 853-862.
141. Costela-Ruiz VJ, Illescas-Montes R, Puerta-Puerta JM, Ruiz C, Melguizo-Rodríguez L. SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. *Cytokine & Growth Factor Reviews*. 2020; 54: 62-75.
142. Ye W, Chen G, Li X, Lan X, Ji C, et al. Dynamic changes of D-dimer and neutrophil-lymphocyte count ratio as prognostic biomarkers in COVID-19. *Respiratory Research*. 2020; 21(1): 169.
143. Yuan S, Jiangm SC, Zhang ZW, Fu YF, Hu J, et al. The Role of Alveolar Edema in COVID-19. *Cells*. 2021; 10(8): 1897.
144. Gouda AS, Mégarbane B. Snake venom-derived bradykinin-potentiating peptides: A promising therapy for COVID-19? *Drug Development Research*. 2021; 82(1): 38-48.
145. Cui Xinyu, Chen Wuyue, Zhou Haoyan, Gong Yuan, Zhu Bowen, et al. Pulmonary Edema in COVID-19 Patients: Mechanisms and Treatment Potential. *Frontiers in Pharmacology*. 2021; 12. 10.3389/fphar.2021.664349.
146. Arthur Y Kim, Rajesh T Gandhi. COVID-19 Management in hospitalized adults, Up To Date. 2022.
147. Fronius M. Treatment of pulmonary edema by ENaC activators/stimulators, *Current Molecular Pharmacology*. 2013; 6(1): 13-27.
148. Aslan A, Aslan C, Zolbanin NM, et al. Acute respiratory distress syndrome in COVID-19: possible mechanisms and therapeutic management. *Pneumonia*. 2021; 13: 14.
149. Adil Mir, Narayanan S Priya, Somanath Payaningal. Is amiloride a promising cardiovascular medication to persist in the COVID-19 crisis?. *Drug discoveries & therapeutics*. 2020; 14: 256-258.
150. Goursaud S, Descamps R, Daubin C, Du Cheyron D, Valette X. Corticosteroid use in selected patients with severe acute respiratory distress syndrome related to COVID-19. *Journal of Infection*. 2020; 81(2): e89-e90.
151. Santos J LF, Zanardi P, Alo V, Rodriguez M, Magdaleno F, et al. Pulmonary Edema in COVID-19 Treated with Furosemide and Negative Fluid Balance (NEGBAL): A Different and Promising Approach. *Journal of Clinical Medicine*. 2021; 10(23): 5599.
152. Abdel Hameid R, Cormet-Boyaka E, Kuebler WM, Uddin M, Berdiev BK. SARS-CoV-2 may hijack GPCR signaling pathways to dysregulate lung ion and fluid transport. *American Journal of Physiology- Lung Cellular and Molecular Physiology*. 2021; 320(3): L430-L435.
153. Sinha M, Zabini D, Guntur D, Nagaraj C, et al. Chloride channels in the lung: Challenges and perspectives for viral infections, pulmonary arterial hypertension, and cystic fibrosis, *Pharmacology & Therapeutics*. 2022; 237: 108249.