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Delayed Composite Mesh Infection: Can *Burkholderia Cepacia* be **Incriminated**?

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Abstract

Delayed mesh infection is a rare complication. In this report, we describe the case of a 68-year-old woman who, underwent cholecystectomy and an eventration on a sub umbilical hernia which required composite mesh implantation with ceolioscopy in 2014. Nine years after the hernia repair, she presented with an infected mesh due to *Burkholderia cepacia*. This infection required medical and surgical treatment with mesh removal in order to cure the patient. The mechanism and etiology of such a late complication are discussed.

Keywords: Composite mesh; Infection; Burkholderia cepacia.

Abbreviations: VHR: Ventral Hernia Repair; PTFE: Polytetrafluoroethylene; BCC: Burkholderia Cepacia Complex; PP: Polypropylene.

Introduction

Delayed mesh infection is a rare complication and the precise mechanism of its development is unknown [1]. Mesh infection, the most devastating mesh-related complication after ventral hernia repair (VHR), may occur in 7% to 10% of patients [2,3]. The most common bacteria associated with prosthetic mesh infection are Staphylococcus aureus (57.7%). Treatment strategies that have been described for mesh infection include complete or partial mesh removal, antimicrobial therapy, and conservative treatment to salvage the mesh [2,4,5]. More recently, management of some patients has shifted to an attempt of conservative treatment (antibiotics, wound management, etc.) to salvage mesh without surgical removal [5]. We report herein a case of composite mesh infection in a 68-year-old patient due to Burkholderia cepacia and after our review by searching PubMed publications till June 2023, our case is the first case of mesh infection secondary to this pathogen.

Case presentation

A 68-year-old woman with a previous medical history of high blood pressure and dyslipidemia, presented with a 15day history of abdominal pain and subocclusif syndrome. She underwent a cholecystectomy in 2013. One year later, she presented an eventration on a sub umbilical hernia which required composite mesh implantation with ceolioscopy in 2014. She remained disease free and asymptomatic from her abdominal incision until 2023 when she developed a subocclusif syndrome and abdominal pain. In january 2023, the patient was admitted with a suspected intra-abdominal cause. On admission, she was apyretic. Abdominal examination revealed a widespread abdominal tenderness, no hepatomegaly, nor splenomegaly. She presented a discrete systolic murmur at the aortic focus. Laboratory investigations revealed elevated C-reactive protein levels at 60 mg/L, a white blood cell count at 4450/mm³, anemia with an hemoglobin level at 9.9 g/dL, and platelets count at

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258000/mm³. Renal and hepatic functions were normal. Blood and urine cultures were collected on admission. Transthoracic echocardiography was normal. Following admission, abdominal Computed Tomography scan (CT) revealed abcess formation with contrast effect regarding the mesh, on the supraumbilical anterior abdominal wall, measuring 11.7 cm transverse diameter x10.5 cm craniocaudal diameter x4.5 cm thick (Figure 1).

The patient underwent CT-guided percutaneous drainage of the abscess. A drain was placed and a sample was taken and sent for bacteriological study. She was started empiric antibiotic therapy based on imipenem 1 gr twice daily, metronidazole 500 mg 3 times daily and pristinamycin 500 mg 2 tablets 3 times daily. Abcess culture grew *Burkholderia cepacia*, which was sensitive to meropnem, minocycline, levofloxacin and cotrimoxazole. The patient had received imipenem combined with levofloxacin 500 mg twice daily and cotrimoxazole 1 pill twice daily. The drainage volume decreased gradually until no fluid was observed. On day 20, the drainage tube was removed.

On day 27 of treatment, the patient had no functional complaints. Laboratory investigations revealed no inflammatory markers. However, abdominal ultrasound showed the persistence of the collection (67x6x84 mm) on the deep side of the mesh.

Following the trial of intravenous antibiotics, a multidisciplinary review recommended that the mesh should be removed (Figure 2). She received metronidazol 500 mg 3 times daily and cotrimoxazole 1 pill twice daily for 2 weeks and levofloxacin 500 mg 2 times daily for 3 weeks. After the removal of the mesh, the patient was reviewed four weeks later with no further infections and a well-healed scar.

Discussion

We report a case of composite mesh infection in a 68-yearold patient due to *Burkholderia cepacia*. We reviewed the literature by searching PubMed publications till June 2023, our case is the first case of mesh infection secondary to this pathogen.

Because of unacceptably high recurrence rates after suturebased VHR, using reinforcing mesh has become a commonly performed and widely accepted procedure [2,6,7]. Although mesh has reduced hernia recurrence rates, it has its own set



Figure 1: Axial section of abdominal computed tomography scan showing abcess formation ($11.7 \times 10.5 \times 4.5 \text{ cm}$) with contrast effectregarding the mesh, on the supraumbilical anterior abdominal wall.

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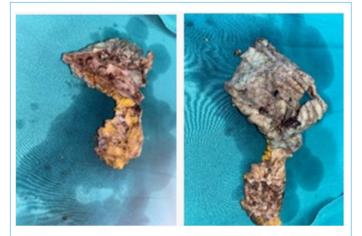


Figure 2: Resected mesh covered with granulation tissue.

of complications. Infection is one of the most devastating complications [2,5]. It is associated with hospital readmission, increased healthcare costs, reoperation, hernia recurrence and impaired quality of life [8].

Mesh infection, the most devastating mesh-related complication after VHR, may occur in 7% to 10% of patients [2,3]. The incidence of mesh infections after incisional hernia repair is about 1% for endoscopic techniques and can be more than 15% in other techniques [9]. Several factors have been identified as being significantly associated with mesh infection and included patient-related factors such as diabetes, obesity, smoking, chronic obstructive pulmonary disease, immunosuppressive treatments [10], factors related to the operating conditions such as strangulated hernia, lack of prophylactic antibiotics at induction and factors related to the prosthesis used [5]. In fact, polytetrafluoroethylene (PTFE) and microporous prostheses are more likely to be associated with prosthesis infection [2,11-14].

Usually, mesh infections occur soon after surgery, within weeks or months at the latest. Delayed infections occuring many years after the abdominal wall mesh insertion are extremely uncommon [14]. One of the mechanisms to be considered is the hematogenous spread of infection and late wound breakdown, which would allow its introduction. Another potential or possible cause is percutaneous drainage of a reactive seroma, which may develop around a longstanding foreign body [14]. According to Jose' Bueno-Lledo et al., the time frame from hernia repair to mesh infection in their series was 10.3 months (range 1 to 29 months) [15]. Indeed, 34.7% of patients presented greater than a year after mesh implantation with a mesh infection [5]. In contrast, A Elfaki et al. reported the longest documented case of mesh infection, occurring over 15 years after mesh insertion [14]. For our patient, the delay was about 9 years after mesh insertion and the mechanism would be probably an hematogenous spread secondary to dental care done two weeks before or an intraoperative revealed very late.

Bacteria typically involved in mesh infection include *Staphylococcus Aureus*, *Staphylococcis coagulase negative* and enteric Gram negative bacteria [8,16]. In the literature, *Staphylococcus aureus* is identified in the majority of prosthetic mesh infections, with a rate estimated in different studies at 81%, 57,7% and 42% [8,17]. However, M. Siebert et al. reported that of the 22 documented mesh infections, 3% were polymicrobial. A total of 54 bacteria were isolated, and the most frequently represented bacteria were *Enterobacteriaceae* (18). For our patient, culture grew *Burkholderia Cepacia*. In fact, the *Burkhold*-

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eria Cepacia Complex (BCC) is a group of non-fermenting and oxidase-positive aerobic Gram-negative bacilli [19-21]. *Burkholderia* causes respiratory infections in immunocompromised patients, especially those with cystic fibrosis and chronic granulomatous disease [22]. It also has been reported in immuno-compromised patients: debilitated elderly people, HIV-positive individuals, cancer patients undergoing chemotherapy [20,23-25]. There have also been reports documenting BCC as being responsible for endocarditis in drug addicts or patients with prosthetic heart valves [26], eye infections following surgery [27] and infections or abscesses of the central nervous system [28]. BCC is also rarely responsible for refractory peritonitis in patients undergoing continuous ambulatory peritoneal dialysis [29,30]. Some cases of splenic and hepatic abcesses were also reported [31,32].

After a careful review of the litterture, we didin't found any case of mesh infection due to *Burkholderia cepacia*. So, we reported the first case of *Burkholderia cepacia* mesh infection. Current treatment for mesh infections is complex due to numerous variables unique to each patient and hernia including patient factors, surgical technique, degree of contamination, mesh type and position, and wound closure [5].

Concerning mesh type, today's prosthetics may be classified broadly into three categories: synthetic, composite, and biologic. Synthetic mesh, such as polypropylene (PP) or polyester, is characterized by high tensile strength and vigorous tissue ingrowth, but is unsuitable for intraabdominal placement because of its tendency to induce bowel adhesions. Composite, or barrier-coated, mesh is a dual-sided prosthetic having a synthetic parietal side to promote a strong repair and a visceral surface that repels tissue ingrowth and decreases adhesion formation [33]. Biologic mesh are derived from human, bovine, and porcine tissue that has been decellularized to leave a collagen matrix and then be incorporated into the host tissue. Biological meshes theoretically generate less of a foreign body response and are more resistant to infection [34].

The current recommended treatment for mesh infection remains complete foreign body removal and antibiotic therapy, including complete removal of mesh and sutures with closure of the fascia [5,9,16,34]. In fact, the initial period of bacterial adhesion can be rapid and reversible. However, subsequent irreversible mesh attachment via bacterial adhesins and production of bacterial biofilm impairs penetration and clearance of bacteria by host immune cells and systemic antibiotics [15,35]. Consequently, when an infection is established, this capsule restricts the penetration of antimicrobial agents into the infected mesh [15], that's why complete removal of mesh is recommended.

More recently, management of some patients has shifted to an attempt of conservative treatment based on antibiotics, wound management and salvage mesh without surgical removal [2,4,5].

In fact, Stremitzer et al. evaluated 31 patients with a mesh infection after VHR and found a mesh salvage rate of 55% with conservative treatment. According to the same study, there was a significant association between the type of mesh graft used and the probability of mesh preservation in case of infection. While conservative therapy led to preservation of 100% of absorbable infected polyglactin/PP meshes, only 20% of infected pure PP meshes and 23% of infected PTFE/PP meshes could be salvaged using conservative means (p<0.0001) [36].

These results are explained in the literature. In fact, in the case of PP meshes, this is most likely due to the pore size inherent in this particular mesh, allowing free flow of bacteria as well as host immune response cells resulting in a more tightly integrated and solid repair [37,38]. On the other hand, PTFE mesh is constructed with very tiny pores, purposefully designed to prevent this free flow of fuid and cells and adhesions. The PTFE layer of this composite type of mesh uniquely creates a diffcult environment to successfully control infection [38,39].

For our patient, she had a composite mesh and several retrospective studies have investigated the incidence of infection after composite mesh repair. In one study, 3.3% of patients undergoing VHR with a composite mesh required mesh explantation because of infection [40].

Greenberg et al reported in their study that eleven patients with composite mesh-related infections, were treated initially with conservative measures. Four patients ultimately required mesh removal. Seven patients, however, were successfully salvaged with conservative methods, including dressing changes, local wound debridement, partial mesh excision, wound vacuum, and antibiotics [38]. None of the patients with a salvaged infected composite mesh developed hernia recurrence in a three-year follow-up. This study stand in contrast to a retrospective review, published by William S Cobb et al. that reported a 10% infection rate among 206 patients who underwent elective composite mesh hernia repair. All but two necessitated mesh removal [3]. Additionally, 95% of heavyweight PPE mesh and all PTFE/PPE meshes have been excised [5]. For our patient mesh excision was due to failure of conservative treatment.

Conclusion

In conclusion, for our patient, the mesh type, the presence of biofilm, failure of conservative treatment (drainage and antibiotics) lead to the mesh excision with a favourable outcome. Each patient must be evaluated and treated on a case-by-case basis. Not all infected composite mesh implants mandate removal. Each patient's presentation must be individualized. Mesh salvage should be considered if possible before condemning patients to mesh explantation and its inherent morbidity.

Declarations

Conflicts of interest: There is no conflicts of interest to declare.

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